AmpliSens® *C.trachomatis / Ureaplasma / M.hominis*-MULTIPRIME-FRT PCR kit

CE 1023

IVD

For Professional Use Only

Instruction Manual

KEY TO SYMBOLS USED

REF	Catalogue number		Caution
LOT	Batch code	Σ	Sufficient for
IVD	*In vitro* diagnostic medical device		Use-by Date
VER	Version		Consult instructions for use
	Temperature limit		Keep away from sunlight
	Manufacturer	NCA	Negative control of amplification
	Date of manufacture	C–	Negative control of extraction
EC REP	Authorized representative in the European Community	C+	Positive control of amplification
		IC	Internal control

1. INTENDED USE

AmpliSens® *C.trachomatis* / *Ureaplasma* / *M.hominis*-MULTIPRIME-FRT PCR kit is an *in vitro* nucleic acid amplification test for simultaneous detection of DNA of *Chlamydia trachomatis, Ureaplasma* (*U.parvum* and *U.urealyticum*) and *Mycoplasma hominis* in the clinical material (urogenital and oropharyngeal swabs; conjunctival discharge; prostate gland secretion; and urine samples) by using real-time hybridization-fluorescence detection.

NOTE: The results of PCR analysis are taken into account in complex diagnostics of disease.

2. PRINCIPLE OF PCR DETECTION

Chlamydia trachomatis, Ureaplasma (U.parvum and *U.urealyticum), Mycoplasma hominis* detection by the multiplex polymerase chain reaction (PCR) is based on the amplification of pathogen genome specific region using specific primers. In the real-time PCR, the amplified product is detected with the use of fluorescent dyes. These dyes are linked to oligonucleotide probes, which bind specifically to the amplified product during thermocycling. The real-time monitoring of fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run.

AmpliSens® *C.trachomatis* / *Ureaplasma* / *M.hominis*-MULTIPRIME-FRT PCR kit is a qualitative test that contains the Internal Control (Internal Control-FL (IC)). It must be used in the extraction procedure in order to control the extraction process of each individual sample and to identify possible reaction inhibition.

AmpliSens® *C.trachomatis* / *Ureaplasma* / *M.hominis*-MULTIPRIME-FRT PCR kit uses "hot-start", which greatly reduces the frequency of nonspecifically primed reactions. "Hot-start" is guaranteed by the separation of nucleotides and Taq-polymerase by using chemically modified polymerase (TaqF). The chemically modified polymerase (TaqF) is contains the system for prevention of contamination by amplicons using the enzyme uracil-DNA-glycosylase (UDG) and deoxyuridine triphosphate. The enzyme UDG recognizes and catalyzes the destruction of the DNA containing deoxyuridine, but has no effect on DNA containing deoxythymidine. Deoxyuridine is absent in the authentic DNA, but is always present in amplicons, because deoxyuridine triphosphate is a part of dNTP mixture in the reagents for the amplification. Due to the deoxyuridine containing contaminating amplicons are sensitive to the destruction by UDG before the DNA-target amplification. So the amplicons cannot be amplified.

The enzyme UDG is thermolabile. It is inactivated by heating at temperature above 50 °C. Therefore, UDG does not destroy the target amplicons which are accumulated during PCR.

The results of amplification are registered in the following fluorescence channels.

Table 1

Channel for fluorophore	FAM	JOE	ROX	Cy5
DNA-target	*Chlamydia trachomatis*	*Ureaplasma* spp.	*Mycoplasma hominis*	Internal Control-FL
Target gene	*cryptic plasmid*	*UreC*	*16s rRNA* gene	genetically engineered construction

3. CONTENT

AmpliSens® *C.trachomatis* / *Ureaplasma* / *M.hominis*-MULTIPRIME-FRT PCR kit is produced in 1 form:

variant FRT-100 F, REF R-B43-F(RG,iQ)-CE.

Variant FRT-100 F includes:

Reagent	Description	Volume, ml	Quantity
PCR-mix-1-FL *C.trachomatis* / *Ureaplasma* / *M.hominis*	clear liquid from colorless to light lilac colour	1.2	1 tube
PCR-mix-2-FRT	colorless clear liquid	0.3	2 tubes
Polymerase (TaqF)	colorless clear liquid	0.03	2 tubes
Positive Control complex (C+)	colorless clear liquid	0.2	1 tube
DNA-buffer	colorless clear liquid	0.5	1 tube
Negative Control (C-)*	colorless clear liquid	1.2	1 tube
Internal Control-FL (IC)**	colorless clear liquid	1.0	1 tube

* must be used in the extraction procedure as Negative Control of Extraction.

** add **10 µl** of **Internal Control-FL (IC)** during the DNA extraction procedure directly to the sample/lysis mixture (see **DNA-sorb-AM** REF K1-12-100-CE).

Variant FRT-100 F is intended for 110 reactions, including controls.

4. ADDITIONAL REQUIREMENTS

- Transport medium.
- DNA extraction kit.
- Disposable powder-free gloves and a laboratory coat.
- Pipettes (adjustable).
- Sterile pipette tips with filter (up to 200 µl).
- Tube racks.
- Vortex mixer.
- Desktop centrifuge with rotor for 2-ml reaction tubes.
- PCR box.
- Real-time instruments (for example, Rotor-Gene 3000/6000 (Corbett Research, Australia); Rotor-Gene Q (QIAGEN, Germany), iCycler iQ5 (Bio-Rad, USA); Mx3000P (Stratagene, USA)).
- Disposable polypropylene PCR tubes (0.1- or 0.2-ml):
  a) 0.2-ml thin-walled PCR tubes with domed caps if a plate-type instrument is used;
  b) 0.2-ml thin-walled PCR tubes with flat caps or strips of four 0.1-ml Rotor-Gene PCR tubes if a rotor-type instrument is used.
- Refrigerator for 2–8 °C.
- Deep-freezer for the temperature from minus 24 to minus 16 °C.
- Reservoir for used tips.

5. GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol filters and use a new tip for every procedure.
- Store all extracted positive material (specimens, controls and amplicons) away from all other reagents and add it to the reaction mix in a distantly separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable protective gloves and laboratory cloths, and protect eyes while samples and reagents handling. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local regulations.
- Samples should be considered potentially infectious and handled in biological cabinet in compliance with appropriate biosafety practices.
- Clean and disinfect all samples or reagents spills using a disinfectant, such as 0.5 % sodium hypochlorite or another suitable disinfectant.
- Avoid inhalation of vapors, samples and reagents contact with the skin, eyes, and mucous membranes. Harmful if swallowed. If these solutions come into contact, rinse the injured area immediately with water and seek medical advice if necessary.
- Safety Data Sheets (SDS) are available on request.
- Use of this product should be limited to personnel trained in DNA amplification techniques.
- Workflow in the laboratory must be one-directional, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where the previous step was performed.

⚠ Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

Not for use in the Russian Federation

6. SAMPLING AND HANDLING

NOTE: Obtaining samples of biological materials for PCR-analysis, transportation, and storage are described in the manufacturer's handbook [1]. It is recommended that this handbook is read before starting work.

AmpliSens® *C.trachomatis / Ureaplasma / M.hominis*-MULTIPRIME-FRT PCR kit is intended for analysis of DNA extracted with DNA extraction kits from the clinical material (urogenital, rectal and pharyngeal swabs; conjunctival discharge; prostate gland secretion; and urine samples).

7. WORKING CONDITIONS

AmpliSens® *C.trachomatis / Ureaplasma / M.hominis*-MULTIPRIME-FRT PCR kit should be used at 18–25 °C.

8. PROTOCOL

8.1. DNA Extraction

It is recommended to use the following nucleic acid extraction kits:

- **DNA-sorb-AM**, REF K1-12-100-CE.
- For other nucleic acid extraction kits see Guidelines [2].

The DNA extraction of each test sample is carried out in the presence of **Internal Control-FL (IC).**

NOTE: Extract the DNA according to the manufacturer's protocol.

8.2. Preparing PCR

8.2.1. Preparing tubes for PCR

The total reaction volume is **25 µl**, the volume of DNA sample is **10 µl**.

1. Thaw the tube with **PCR-mix-2-FRT.** Mix the tubes with **PCR-mix-1-FL *C.trachomatis / Ureaplasma / M.hominis,* PCR-mix-2-FRT, polymerase (TaqF).** Sediment the drops by short centrifugation (1-2 s) using vortex.
   Prepare the required number of the tubes for amplification of DNA from clinical and control samples.
2. For N reactions (including 2 controls), add to a new tube:
   10*(N+1) µl of PCR-mix-1-FL *C.trachomatis / Ureaplasma / M.hominis*,
   5.0*(N+1) µl of PCR-mix-2-FRT
   0.5*(N+1) µl of polymerase (TaqF).
   Vortex the tube, then centrifuge briefly. Transfer **15 µl** of the prepared mixture into each tube.
3. Add **10 µl** of **DNA samples** obtained at the DNA extraction stage into prepared tubes.
4. Carry out the control amplification reactions:
   NCA – Add **10 µl** of **DNA-buffer** to the tube labeled NCA (Negative Control of Amplification).
   C+ – Add **10 µl** of **Positive Control complex** to the tube labeled C+ (Positive Control of Amplification).
   C– – Add **10 µl** of **the sample extracted from the Negative Control reagent** to the tube labeled C– (Negative control of Extraction).

8.2.2. Amplification

1. Create a temperature profile on your instrument as follows:

Table 2

AmpliSens-1 RG amplification program

Step	Rotor-type instruments[1]			Plate-type instruments[2]		
	Temperature, °C	Time	Cycles	Temperature, °C	Time	Cycles
1	95	15 min	1	95	15 min	1
2	95	5 s	5	95	5 s	5
	60	20 s		60	20 s	
	72	15 s		72	15 s	
3	95	5 s	40	95	5 s	40
	60	20 s (fluorescence detection)		60	30 s (fluorescence detection)	
	72	15 s		72	15 s	

Fluorescence is detected in FAM/Green, JOE/Yellow, ROX/Orange, and Cy5/Red fluorometer channels. Fluorescent signal is detected in the channels for the FAM, JOE, ROX and Cy5 fluorophores (other channels are enabled if several tests are simultaneously carried out in a single run).

2. Adjust the fluorescence channel sensitivity according to the *Important Product Information Bulletin* and Guidelines [2].
3. Insert tubes into the reaction module of the device.
4. Run the amplification program with fluorescence detection.
5. Analyze results after the amplification program is completed.

9. DATA ANALYSIS

Analysis of results is performed by the software of the real-time PCR instrument used by measuring fluorescence signal accumulation in four channels:

- The signal of the *Chlamydia trachomatis* DNA amplification product is detected in the channel for the FAM fluorophore.
- The signal of the *Ureaplasma* spp. *(U.parvum* and *U.urealyticum)* DNA amplification product is detected in the channel for the JOE fluorophore.
- The signal of the *Mycoplasma hominis* DNA amplification product is detected in the channel for the ROX fluorophore.
- The signal of the IC DNA amplification product is detected in the channel for the Cy5 fluorophore.

Results are interpreted by the crossing (or not-crossing) the fluorescence curve with the threshold line set at the specific level that corresponds to the presence (or absence) of a *Ct* value of the DNA sample in the corresponding column of the results grid.

Principle of interpretation is the following:

- ***Chlamydia trachomatis*** DNA is **detected** if the *Ct* value is determined in the results grid in the channel for the FAM fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
- ***Ureaplasma* spp. *(U.parvum* and *U.urealyticum)*** DNA is **detected** if the *Ct* value is determined in the results grid in the channel for the JOE fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
- ***Mycoplasma hominis*** DNA is **detected** if the *Ct* value is determined in the results grid in the channel for the ROX fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
- ***Chlamydia trachomatis, Ureaplasma spp.* and *Mycoplasma hominis*** DNA are **not detected** in a sample if the *Ct* value is not determined (absent) in the channels for FAM, JOE and ROX fluorophores (fluorescence curve does not cross the threshold line), whereas the *Ct* value determined in the channel for the Cy5 fluorophore is less than the boundary *Ct* value specified in the *Important Product Information Bulletin*.
- The result is **invalid** if the *Ct* value is not determined (absent) in the channel for Cy5 fluorophore and in the channels for the FAM, JOE and ROX fluorophores. In such case, the PCR analysis of corresponding sample should be repeated.

NOTE: Boundary *Ct* values are specified in the *Important Product Information Bulletin* enclosed to the PCR kit. See also Guidelines [2].

The result of the analysis is considered reliable only if the results obtained for Positive and Negative Controls of amplification as well as for the Negative Control of extraction are correct (see Table 3).

Table 3

Results for controls

Control	Stage for control	*Ct* value in the channel for fluorophore	
		FAM, JOE, ROX	Cy5
C–	DNA extraction	Absent	<boundary value
NCA	PCR	Absent	Absent
C+	PCR	<boundary value	<boundary value

10. TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

1. If the *Ct* value determined for the Positive Control of Amplification (C+) in the channels for the FAM and/or JOE and/or ROX fluorophores is greater than the boundary *Ct* value or absent, the amplification should be repeated for all samples in which the *Ct* value was absent in the channels for the FAM and/or JOE and/or ROX fluorophores.
2. If the *Ct* value is determined for the Negative Control of Amplification (NCA) and/or Negative Control of Extraction (C–) in the channels for the FAM and/or JOE and/or ROX fluorophores, the PCR analysis should be repeated starting from the DNA extraction stage for all samples in which the *Ct* value was determined in the channels for the FAM and/or JOE and/or ROX fluorophores.

If you have any further questions or if encounter problems, please contact our Authorized representative in the European Community.

11. TRANSPORTATION

AmpliSens® *C.trachomatis / Ureaplasma / M.hominis*-MULTIPRIME-FRT PCR kit should be transported at 2–8 ºC for no longer than 5 days.

12. STABILITY AND STORAGE

All components of the **AmpliSens® *C.trachomatis / Ureaplasma / M.hominis*-MULTIPRIME-FRT** PCR kit are to be stored at 2–8 ºC when not in use (except for Polymerase (TaqF) and PCR-mix-2-FRT. All components of the **AmpliSens® *C.trachomatis / Ureaplasma / M.hominis*-MULTIPRIME-FRT** PCR kit are stable until the expiry date stated on the label. The shelf life of reagents before and after the first use is the same, unless otherwise stated.

NOTE: Polymerase (TaqF) and PCR-mix-2-FRT are to be stored at the temperature from minus 24 to minus 16 °C.

NOTE: PCR-mix-1-FL *C.trachomatis / Ureaplasma / M.hominis* is to be kept away from light.

[1] For example, Rotor-Gene 3000, Rotor-Gene 6000, Rotor-Gene Q or equivalent
[2] For example, iCycler, iQ5, Mx3000P, Mx3000, DT-96 or equivalent.

13. SPECIFICATIONS

13.1. Sensitivity

Clinical material	Nucleic acid extraction kit	PCR kit	Microorganism	Sensitivity, GE/ml[3]
Urogenital swabs [4]	**DNA-sorb-AM**	variant FRT-100 F	*Chlamydia trachomatis*	$5x10^2$
			Ureaplasma spp.	10^3
			Mycoplasma hominis	10^3
Urine [5]	**DNA-sorb-AM**	variant FRT-100 F	*Chlamydia trachomatis*	10^3
			Ureaplasma spp.	$2x10^3$
			Mycoplasma hominis	$2x10^3$

NOTE: The analytical sensitivity of each microorganism does not change even if two other microorganisms are present at high concentrations.

13.2. Specificity

The analytical specificity of **AmpliSens® *C.trachomatis* / *Ureaplasma* / *M.hominis*-MULTIPRIME-FRT** PCR kit is ensured by the selection of specific primers and probes as well as stringent reaction conditions. The primers and probes were checked for possible homologies to all sequences published in gene banks by sequence comparison analysis.
Nonspecific responses were absent while testing human DNA samples and DNA samples of microorganisms: *Gardnerella vaginalis, Lactobacillus* spp., *Escherichia coli, Staphylococcus* spp., *Streptococcus* spp., *Candida albicans, Ureaplasma urealyticum, Ureaplasma parvum, Mycoplasma hominis, Chlamydia trachomatis, Mycoplasma genitalium, Neisseria* spp., *Neisseria gonorrhoeae, Trichomonas vaginalis, Treponema pallidum, Toxoplasma gondii, HSV* types 1 and 2, *CMV,* and *HPV*.
The clinical specificity of **AmpliSens® *C.trachomatis* / *Ureaplasma* / *M.hominis*-MULTIPRIME-FRT** PCR kit was confirmed in laboratory clinical trials.

14. REFERENCES

1. Handbook “Sampling, Transportation, and Storage of Clinical Material for PCR diagnostics”, developed by Federal Budget Institute of Science “Central Research Institute for Epidemiology” of Federal Service for Surveillance on Consumers’ Rights Protection and Human Well-Being.
2. Guidelines “Real-Time PCR Detection of STIs and Other Reproductive Tract Infections”, developed by Federal Budget Institute of Science “Central Research Institute for Epidemiology”.

15. QUALITY CONTROL

In compliance with Federal Budget Institute of Science “Central Research Institute for Epidemiology” ISO 13485-Certified Quality Management System, each lot of the **AmpliSens® *C.trachomatis* / *Ureaplasma* / *M.hominis*-MULTIPRIME-FRT** PCR kit has been tested against predetermined specifications to ensure consistent product quality.

List of Changes Made in the Instruction Manual

VER	Location of changes	Essence of changes
23.06.11 RT	Cover page, text	The name of Institute was changed to Federal Budget Institute of Science “Central Research Institute for Epidemiology”
24.09.15 ME	Text	Corrections according to the template
	1. Intended use	The clinical material was specified
	6. Sampling and handling	
	3. Content, Footer	REF R-B43(iQ)-CE was deleted
	9. Data analysis	The sections were rewritten
	10. Troubleshooting	
25.12.17 PM	3. Content	The color of the reagent was specified
05.12.18 EM	2. Principle of PCR detection	The table with targets and the information about the enzyme UDG were added
	Through the text	The text formatting was changed
27.02.20 PM	Footer	The phrase “Not for use in the Russian Federation” was added
26.10.20 KK	Footer, 3. Content	The information about variant FRT REF R-B43-(RG)-CE was deleted
01.03.21 MA	—	The name, address and contact information for Authorized representative in the European Community was changed

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[3] The quantity of genome equivalents of microorganism per 1 ml of the clinical sample placed into the transport medium.
[4] Cervical and urethral swabs are to be placed into **Transport Medium for Swabs** (REF 956-CE, REF 987-CE) or **Transport Medium with Mucolytic Agent** (REF 952-CE, REF 953-CE).
[5] Pretreatment is required.